International Rules for Seed Testing
2019
Validated Seed Health Testing Methods

7-014: Detection of *Parastagonospora nodorum* in *Triticum aestivum* (wheat) seed

Including changes and editorial corrections adopted at the Ordinary General Meeting 2018, Sapporo, Japan

Effective from 1 January 2019
Chapter 7: Validated Seed Health Testing Methods

Validation reports

See References. Copies are available by e-mail from the ISTA Secretariat at ista.office@ista.ch.

Please send comments, suggestions or reports of problems relating to this method to the ISTA Seed Health Committee, c/o ISTA Secretariat.

Disclaimer

Whilst ISTA has taken care to ensure the accuracy of the methods and information described in this method description, ISTA shall not be liable for any loss or damage, etc. resulting from the use of this method.

Safety precautions

Ensure you are familiar with hazard data and take appropriate safety precautions, especially during weighing out of ingredients. It is assumed that persons carrying out this test are in a laboratory suitable for carrying out microbiological procedures and familiar with the principles of Good Laboratory Practice, Good Microbiological Practice, and aseptic techniques. Dispose of all waste materials in an appropriate way (e.g. autoclaving, disinfection) and in accordance with local health, environmental and safety regulations.

Note on the use of the translations

The electronic version of the International Rules for Seed Testing includes the English, French, German and Spanish versions. If there are any questions on interpretation of the ISTA Rules, the English version is the definitive version.

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**7-014: Detection of *Parastagonospora nodorum* in *Triticum aestivum* (wheat) seed**

**Host:** *Triticum aestivum* L.


**Prepared by:** ISTA-PDC Method Validation Sub-committee

**Authors:** ISTA-PDC Method Validation Sub-committee

**Revision history**

Version 1.0, 2001-11-20
Revised 2001-11-20, J. Sheppard
Reprinted with editorial changes 2003
Version 1.1, 2008-01-01: Treated seed revised; Reporting results revised
Version 1.2, 2010-01-01: Footnote 2 added to malt agar ingredients
Version 1.3, 2014-01-01: Streptomycin sulphate added to malt agar; addition of positive control; pathogen renamed *Stagonospora*; reference culture mandatory
Version 1.4, 2017-01-01: Reporting results revised
Version 1.5, 2018-01-01: Changes to the taxonomic names of fungi

**Background**

This method was originally published in the *ISTA Handbook of Seed Health Testing* in November 1964 as S.3. No. 19 and revised in 1984 by M. Kietreiber, Bundesanstalt für Pflanzenbau, Wien, Austria. The method was incorporated into the newly revised *Annexe to Chapter 7* in 2002 from the 1999 edition of the ISTA Rules. The method was reviewed by the ISTA-Seed Health Committee in 2006 (Cockerell & Koenraadt, 2007) with the recommendation to accept for a further five years.

**Treated seed**

This method has not been validated for the determination of *Parastagonospora nodorum* on treated seed. Seed treatments may affect the performance of the method. (Definition of treatment: any process, physical, biological or chemical, to which a seed lot is subjected, including seed coatings. See 7.2.3.)

**Materials**

**Reference material:** reference cultures or other appropriate material

**Media:** malt agar or potato dextrose agar with streptomycin sulphate

**Sodium hypochlorite solution:** (1 % available chlorine)

**Petri dishes:** when sowing density is given by a number of seeds per Petri dish, a diameter of 90 mm is assumed

**Incubator:** capable of operating in the range 20 ±2 °C

**Sample preparation**

The test is carried out on a working sample of 400 seeds as described in Section 7.4.1 of the ISTA Rules.

**Methods**

1. Pretreatment: 10 min in 1 % (available chlorine) sodium hypochlorite.
2. Plating:
   2.1 Aseptically place a maximum of 10 seeds, evenly spaced, onto the agar surface of each malt agar or potato dextrose agar plate.
   2.2 Positive control (reference material): Aseptically place seeds in an appropriate number of plates to obtain the reference culture, or plate a reference culture on media. The number of plates required will depend on the level of contamination of the positive control seed lot.
3. Incubation: 7 days at 20 °C in darkness.
4. Examination: After 7 days examine each seed by naked eye for slow-growing circular colonies of dense white or cream mycelium that often covers infected seeds. The reverse of the colony is yellow/brown becoming darker with age. Compare with positive control.
Chapter 7: Validated Seed Health Testing Methods

General methods

Checking tolerances: Tolerances provide a means of assessing whether or not the variation in results within or between tests are sufficiently wide as to raise doubts about the accuracy of the results. Suitable tolerances, which can be applied to most direct seed health tests, can be found in Table 5B Part 1 of Chapter 5 of the ISTA Rules, or Table G1 in Miles (1963).

Reporting results: The result of a seed health test should indicate the scientific name of the pathogen detected and the test method used. When reported on an ISTA Certificate, results are entered under ‘Other Determinations’.

The report must indicate the number of seeds tested. In the case of a negative result (pathogen not detected), the results must be reported as ‘not detected’.

In the case of a positive result, the report must indicate the percentage of infected seeds.

Quality assurance

Critical control points (CCP)

Where the wording of the original Working Sheet suggests that an action is critical, this has been marked with CCP.

Media and solutions

Sodium hypochlorite solution

Sodium hypochlorite for pretreatment of seed can be prepared from commercial bleach diluted to 1 % available chlorine. The concentration of chlorine in commercial bleach varies considerably. Use the formula:

\[ V_{stock} = V_{final} \times C_{final} / C_{stock} \]

(where \( V = \) volume and \( C = \% \) available chlorine) to calculate the volume of commercial bleach stock solution required to prepare sodium hypochlorite solutions for use in seed pretreatment.

To prepare a 1 L solution of sodium hypochlorite containing 1 % chlorine from a stock of commercial bleach containing 12 % available chlorine:

\[ V_{stock} = V_{final} \times C_{final} / C_{stock} \]

\[ V_{stock} = 1 \times 1/12 = 0.083 \]

Thus add 83 mL of the 12 % stock to 917 mL water.

Malt agar

Malt agar\(^1\): according to manufacturer's instructions

Distilled/deionised water: 1000 mL

Streptomycin sulphate\(^2\): may be used between 50 and 100 ppm, depending on the level of saprophytic bacterial contamination commonly encountered

1 CCP: Malt agar constituents should be equivalent to those of the following manufacturers: BD, USA or Oxoid, UK.

\(^2\) Added after autoclaving

Preparation

1. Weigh out ingredients into a suitable autoclavable container.
2. Add 1000 mL of distilled/deionised water.
3. Dissolve powdered malt agar in distilled/deionised water by stirring.
4. Autoclave at 121 °C and 15 psi for 15 min.
5. Allow agar to cool to approx. 50 °C, and add streptomycin sulphate dissolved in water.
6. Pour 15–22 mL of molten agar into 90 mm Petri plates and allow to solidify before use.

Storage

Prepared plates may be stored at 4 °C for up to 6 weeks.
Potato dextrose agar (PDA)

Potato dextrose agar\(^1\): according to manufacturer's instructions

Distilled/deionised water: 1000 mL

Streptomycin sulphate\(^2\): may be used between 50 and 100 ppm, depending on the level of saprophytic bacterial contamination commonly encountered

\(^1\) CCP: PDA constituents should be equivalent to those of the following manufacturers. BD, USA or Oxoid, UK.

\(^2\) Added after autoclaving

Preparation

1. Weigh out ingredients into a suitable autoclavable container.
2. Add 1000 mL of distilled/deionised water.
3. Dissolve powdered PDA in distilled/deionised water by stirring.
4. Autoclave at 15 psi and 121 °C for 15 min.
5. Allow agar to cool to approx. 50 °C, and add streptomycin sulphate dissolved in water.
6. Pour 15–22 mL of molten agar into 90 mm Petri plates and allow to solidify before use.

Storage

Prepared plates may be stored at 4 °C for up to 6 weeks.
References

The following references are extracted from the *ISTA Handbook of Seed Health Testing*, Working Sheet No. 19, M. Kietreiber, 1984.


Klitgård, K. & Jørgensen, J. (1973). Sammenhængen mellem spireevnen bestemt i laboratorium og mark ved forekomst af *Septoria nodorum* i udsæd af vinterhvede (The correlation between the germination percentage determined in the laboratory and the field with samples of seeds of winter wheat infected with *Septoria nodorum*). *Statsfrokontrollens beretning*, 102, 85–93, København.


Validation references

**Studied in international comparative testing:** 1959, 1961, 1962, 1964 and 1979–81

Using potato dextrose in darkness, Hewett (1975) found a correlation coefficient of 0.95 between counts in the laboratory and the number of diseased seedlings in the field. Comparative tests organized by the ISTA Plant Disease Committee gave reasonable agreement between stations (Rennie, 1982).