Dormancy Breaking for Germination Testing

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Professional Information & "Disclaimer"



- Accredited seed analyst by SCST (non-govt. org) and AOSA (govt. org) for several years
- Laboratory experience testing a wide range of species (e.g., agricultural, forage, turf, vegetable, flower, tree, shrub, native species, etc.)
- Worked in govt., seed cert., commercial, and international company (ISTA accredited) seed laboratories
- Experience using the ISTA Rules, AOSA Rules, USDA Rules, and Canadian Rules for testing seeds

Conducted science-based validation studies using ISTA protocols

- Serve as the Vice-Chair of the ISTA Germination TCOM
- Serve as the SCST Co-Chair of the AOSA/SCST Germination and
 - **Dormancy Sub-Committee**
- Not a seed physiologists...not a PhD...but I have friends that are...





But just like some of us, some seeds like to "sleep late" and are slow to "wake"...that is...to <u>GERMINATE</u>!!! In the world of seeds, this is commonly refereed to as "<u>DORMANCY</u>".

A common question is...." Just how do you '<u>wake up</u>' a seed?"





- Brief overview of what dormancy is/is not
- Rudimentary requirements for germination
- Basic types of dormancy
- ISTA Handbook on Seedling Evaluation Introduction
- Purpose of Dormancy Breaking Procedures
- ISTA Rules Dormancy Breaking Procedures
- Function of Dormancy Breaking Procedures
- Reporting on ISTA Orange Certificate (OIC)



Seed Dormancy -

Refers to a situation in which viable seeds fail to germinate, even when provided with normally favorable conditions that initiate germination.

Note: Seeds must be **imbibed** and determined to be **viable** to be classified as "**dormant**". Also, "**dormant**" is <u>NOT</u> the same as "**hardseededness**" or "**state of quiescence**"...because in these two conditions seeds have not imbibed moisture.

Basic conditions typically required for germination:

- 1) Appropriate <u>moisture</u>
- 2) Appropriate <u>temperature</u>
- 3) Adequate <u>air</u> exchange (exceptions e.g., Oryza sativa/Rice)
- 4) Exposure to <u>light</u> (natural or artificial) required by some species











Primary/Enforced Dormancy –

These types of dormancies delay germination of the seed.

- Physical dormancy caused by the seed coat
- Mechanical dormancy caused by the seed coat
- Chemical dormancy
- Morphological dormancy
- Physiological or Intermediate dormancy
- Deep dormancy
- Double dormancy



Primary/Enforced Dormancy –

- Seed coat physical dormancy prevents the seed from absorbing water
- Seed coat mechanical dormancy when the seed coat prevents the embryo from expanding and emerging from the seed coat
- Chemical dormancy is when internal chemicals inhibit/prevent the seed from germinating



Primary/Enforced Dormancy –

- Morphological dormancy is when the embryo is immature when the seed separates from the mother plant
- Physiological or Intermediate dormancy occurs because a physiological condition is not being met
- Deep dormancy is when seeds remain dormant until favorable conditions are met
- Double dormancy is the combination of any of the previously mentioned factors



Secondary/Induced Dormancy –

- Caused by temperature extremes, too hot or too cold
- Caused by prolonged darkness or prolonged light
- Caused by water stress, too much or not enough
- Caused by lack of oxygen/gas exchange

Seed Dormancy Information: ISTA Hndbk on Seedling Evaluation

ISTA Handbook on Seedling Evaluation provides...

- Essential information on seed physiology (Dormancy - Sec. 2.9)
- Conditions for carrying out germination tests in laboratories
- Gives educational and illustrated guidance \bullet for the evaluation of seedlings for a large number of species
- Supplements and provides additional information on the methods described in the ISTA Rules





ISTA Publications https://www.seedtest.org/en/handbooks-calibration-samples/seedling-evaluation-4th-edition-2018-product-1016.html



lection 2: Germinal



Chapter 5: The germination test

5.1 Object

"The **object of the germination test** is to determine the **germination potential** of a seed lot, which can then in turn be used to compare the quality of different lots and also estimate the field planting value..."

5.2.1 Germination

Germination of a seed in an ISTA test is the emergence and development of the seedling to a stage where the aspect of its essential structures indicates whether or not it is able to develop further into a satisfactory plant under favourable conditions in the field."



Chapter 5: The germination test

5.6.3 Procedures for promoting germination of dormant seed "...If **dormancy** is suspected, **more complete germination** may be obtained by **retesting** after one or a combination of **dormancy-breaking procedures**...and all other procedures listed in 5.6.3.1, 5.6.3.2, and 5.6.3.3 can be used for any species without restriction..."



ISTA Rules - Procedures for Breaking Dormancy

Dormancy breaking procedures listed in 5.6.3.1

- Prechilling
- Preheating
- Prestorage*

 (*This is actually "State of Quiescence", seeds are not imbibed.)
- Light (Minimum of 8 hours per 24-hour period.)
- Sealed polyethylene envelopes/resealable plastic bags (Increases ethylene gas and carbon dioxide.)
- Gibberellic acid (GA3) solution
- Potassium nitrate (KNO3) solution
- Acid scarification
- Mechanical scarification



Prechilling: The replicates for germination are placed in contact with the moist substrate and kept at a low temperature for an initial period before they are moved to

International Rules for Seed Testing

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SECONESTING

ISTA Rules - Procedures for Breaking Dormancy

Dormancy breaking procedures NOT listed in 5.6.3.1, but found in Table5A

- Prewashing
- Presoaking



- Removal of enclosing structures (e.g., pericarp, pod)
- Incubation in moist substrate
- Alternating temperatures during the germination test period (*e.g., 20 ⇔30C*)
- Ethylene gas* (*No procedure provided.*)

(*Ethylene mentioned in ISTA Hndk. on Sdlg. Eval. 2.9.1.3.5 for *Helianthus/Oryza* but no procedure given.)





Treatments BEFORE the germination test

- Prestorage (Quiescence)
- Prechilling
- Incubation in moist substrate
- Preheating
- Prewashing
- Presoaking
- Removal of enclosing structures
- Mechanical scarification
- Acid scarification







Treatments **DURING** the germination test

- Light
- Alternating temperatures during the germination test period (*e.g., 20 ⇔30C*)
- Sealed **polythene/polyethylene envelopes** (*e.g., resealable plastic bags*)
- **Gibberellic acid** (GA3) solution applied to germination test substrate
- Potassium nitrate (KNO3) solution applied to germination test substrate



In OC.

ISTA Rules - Table 5A Dormancy Breaking Info



Dormancy-breaking methods are found in the *"Recommendations for breaking dormancy"* column in Table 5A Part 1, 2, and 3 of the ISTA Rules.

Species	Substrate	Temperature* (°C)	First count (d)	Final count (#)	Recommendations for breaking dormancy	Additional directions	Additional advice
1	2	3	4	5	6	7	8
Abelmoschus esculentus	TP; BP; S	20⇔30	-	21	-	-	-
Achillea millefolium	TP	20⇔30	5	14	-	-	-
Aeschynomene americana	TP	20@35:20@30	4	14	-	-	-
Agropyron cristatum	TP	20->30; 15=25	5	14	KNO _a ; prechill	-	-
Agropyron desertorum	TP	20@30; 15@25	5	14	KNO ₃ ; prechill	-	-
Agrostis canina	TP	20∞30; 15∞25;	7	21	KNO ₃ ; prechill	-	-

Table 5A Part 2. Detailed methods for germination tests: tree and shrub seeds (continued)

Table 5A Part 1. Detailed methods for germination tests: agricultural and vegetable seeds

Species	Substrate	Temperature* (C)	First count (d)	Final count (d)	Recommendation for breaking dormancy	Additional directions	Additional advice
1	2	3	4	5	6	7	8
Acer tebrum, Acer saccharioum	S; (TP)	20	7	21	-	-	-
Aesculus hippocastenum	TS; (S)	20⇔30; (20)	7	21	Soak in water for 48 h; cut off scar end of seed. Do not remove testa from sown por- tion. Fresh nuts may require prechill	-	-

Chapter 5: The germination test

Table 5A Part 3. Detailed methods for germination tests: flower, spice, herb and medicinal species

Species	Substrate	Temperature* (°C)	First count (d)	Final count (d)	Recommendations for breaking dormancy
1	2	3	4	5	6
Abutilon ×hybridum	TP; BP	20⇔30; 20	5–7	21	-
Achillea clavennae	TP; BP	20⇔30; 20	5	14	Light
Achillea filipendulina	TP; BP	20⇔30; 20	5	14	Light
Achillea ptarmica	TP; BP	20⇔30; 20	5	14	Light
Achillea umbellata	TP; BP	20⇔30; 20	5	14	Light



- Simulates cooler/cold winter conditions that often break dormancy
- Artificially decrease inhibitors and increase promoters inside of seeds
- Disrupt the seed coat to allow for imbibition and gaseous exchange
- Stimulate germination with exposure of the seed to light
- Simulate diurnal fluctuation of temperatures of day and night cycle
- Stimulate internal activities of the seed to encourage germination
- Increase natural concentrations of ethylene and carbon dioxide
- Leach away germination inhibitors in the seed
- Removal of seed structures that physically inhibit germination

(More details in the ISTA Handbook on Seedling Evaluation Sec. 2.9)



*****Caution: Always be aware!!!*****

Adverse germination **test conditions** can cause **secondary dormancy** (e.g., media too dry/too wet, inadequate gaseous exchange, no light during germination test).

This phenomenon can cause **inaccurate test results** for **all** or **some** of the **replicates**. This induced dormancy can also be **greatly inconsistent** and can vary by species, seed lot, age of seed lot, test method, test replicates, etc.

Just because a seed does not germinate during the germination test does not automatically mean it is dead!!!



Chapter 6: Biochemical test for viability: the topographical tetrazolium test **Basic Process:**

6.1 Object

The objects of biochemical tests are:

- a. To make a quick estimate of the viability of seed samples in general and those showing dormancy in particular.
- b. In the case of particular samples which at the end of a germination test reveal a high percentage of dormant seeds (5.6.5), to determine the viability of individual dormant seeds or the viability of a working sample.



The **tetrazolium test** is the method typically used to determine the viability of ungerminated seeds, remaining at the end of the germination test.

Imbibed seeds are carefully **cut**, allowing the embryo to be exposed to the **tetrazolium solution**. Respiring living seed tissue will **stain red** in color.

After an appropriate time soaking in the tetrazolium solution, the staining patterns and soundness of the tissue are **evaluated**. Seeds are then classified as "**viable**" or "**non-viable**". Viable seeds are reported on the ISTA certificate as "fresh seeds" and non-viable seeds are reported as "dead seeds".

Biochemical Viability Determination of Ungerminated Seeds



Example for Cereals:

Cut imbibed seeds remaining at the end of the germination test



- a. Longitudinal bisection cut through embryo
- b. Transverse cut above embryo



Figure 6.3b. Evaluation guide for cereals: non-viable seeds.

A Figures are representative for *Triticum,* ×*Triticosecale, Secale, Hordeum* and *Avena* when prepared by bisection or bisected for evaluation

Figure 6.3a. Evaluation guide for cereals: viable seeds.

The figures in column I are all completely stained and viable. Columns **II–V** show the maximum area of unstained, flaccid or necrotic tissue permitted in viable seeds.

- A The figures are representative for *Triticum*, ×*Triticosecale*, *Secale*, *Hordeum* and *Avena* when prepared by bisection or bisected for evaluation
- B Avena prepared by transverse cutting



Reporting info on the OIC



1) ISTA Rules 1.5.2.6 and 5.6.3 REQUIRES any "**special treatment and/or methods used to promote germination**" MUST be stated on the OIC under "**Other determinations**". This informs the person reading the OIC if any methods were used to break dormancy.

2) If there are **5% or more suspected fresh seeds** found in the germination test, their **viability must be determined** and reported on the OIC. There is a required process to determine the viability of the ungerminated seeds to assess if they are "fresh seeds" or "dead seeds".

3) Labs have the option to conduct "**Parallel Tests**", ISTA Rules 5.2.3, with one test using **germination promoting procedures/methods** and one test **without**. Both test results can be reported on the OIC.

The important issue is that the percentage of fresh/dormant seed is determined in a seed lot before it is sold and planted. This info should be provided to the end user that is going to sow/plant this seed lot. To most growers, **"fresh seeds" are considered "dead seeds"** because the seed will typically not germinate within the grower's desired time frame. There will be no uniform emergence of the seedlings when the seed lot has a significant percent of fresh/dormant seeds.



Reporting info on the OIC



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MUST report dormancy breaking procedures used in "Other determinations" and the percent "Fresh seeds" determined by appropriate procedures.

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- ISTA Rules
- ISTA Handbook on Seedling Evaluations
- Seed Dormancy Drew Mutlick, Erin Milligan, and Mark Mullin

Link https://www.powershow.com/view/b54c3-NjU5Y/Seed Dormancy powerpoint ppt presentation

Thank you!

04-10 May

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