



International Rules for Seed Testing
Annexe to Chapter 7: Seed Health Testing Methods



7-012: Detection of *Alternaria padwickii* on *Oryza sativa* (Rice)

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DISCLAIMER: whilst ISTA has taken care to ensure the accuracy of the methods and information described in this method description ISTA shall not be liable for any loss damage etc., resulting from the use of this method.

- Crop:** *Oryza sativa* (Rice)
- Pathogen:** *Alternaria padwickii* (Ganguly) M.B. Ellis syn. *Trichoconis padwickii* Ganguly *Trichoconiella padwickii* (Ganguly) Jain
- Prepared by:** ISTA-PDC Method Validation Sub-committee
- Revision History:** Version 1.0 July 13, 2000
Revised 20.11.2001 J. Sheppard, V. Cockerell
Reprinted 2003
Version 1.1 2008-01-01
“Treated seed” revised; “Reporting results” revised
- Submitted by:** ISTA-PDC Method Validation Sub-committee

Background

This method was originally published in the ISTA Handbook of Seed Health Testing in November 1964 as S.3. No. 13. The method was incorporated into the newly revised Annexe to Chapter 7 in 2002 from the 1999 edition of the ISTA Rules. The method was reviewed by the ISTA-Seed Health Committee in 2006 (Cockerell & Koenraad, 2007) with the recommendation to accept for a further five years.

Studied in International Comparative Testing: 1963, 1964

Safety Precautions

Ensure you are familiar with hazard data and take appropriate safety precautions, especially during preparation of media, autoclaving and weighing out of ingredients. It is assumed that this procedure is being carried out in microbiological laboratory by persons familiar with the principles of Good Laboratory Practice, Good Microbiological Practice, and aseptic technique. Dispose of all waste materials in an appropriate way (e.g. autoclave, disinfect) and in accordance with local safety regulations.

Treated Seed

This method has not been validated for the determination of *Alternaria padwickii* on treated seed. Seed treatments may affect the performance of the method.

(Definition of treatment: any process, physical, biological or chemical, to which a seed lot is subjected, including seed coatings. See 7.2.3)

Materials

- Reference Material** - The use of reference cultures or other appropriate material is recommended when ever possible.
- Media** - Blotters (filter paper)
- Petri dishes** - When sowing density is given by a number of seeds per Petri dish, a diameter of 90 mm is assumed.
- Incubator** - Capable of operating in the range 22 ± 2 °C. To stimulate sporulation, alternating 12 h periods of darkness and near-ultraviolet light (NUV) during incubation are recommended. The recommended source is the *black light* fluorescent lamp (peak at 360 nm) but daylight fluorescent tubes are satisfactory.

Sample Preparation

The test is carried out on a working sample of 400 seeds as described in Section 7.4.1 of the International Rules for Seed Testing.

Method

1. *Pretreatment*
None
2. *Blotter*
On water-soaked blotters in Petri dishes. Place 25 seeds in each dish.
3. *Incubation*
7 days at 22 °C, preferably in NUV in 12 h light/12 h dark cycles.
4. *Examination*
Examine each seed at $\times 12$ -50 magnification for conidia of *A. padwickii*. Conidia are fusiform, first sub-hyaline, later straw coloured to golden brown with long beaks, borne singly either on short conidiophores arising straight from the seed coat or on conidiophores within white-grey, fluffy, aerial mycelium. In doubtful cases confirmation

may be made by examining conidia at $\times 200$ magnification. Conidia are 3-5 septate, often constricted at the septa with characteristic conical basal cells and long beaks, $95\text{--}170\ \mu\text{m} \times 11\text{--}20\ \mu\text{m}$. Infected seeds or seedlings are often surrounded by a characteristic pink stain which diffuses out into the blotter and becomes more intense after 7 days.

General Methods (common to many test procedures)

1. Checking tolerances

Tolerances provide a means of assessing whether or not the variation in result within or between tests is sufficiently wide as to raise doubts about the accuracy of the results. Suitable tolerances, which can be applied to most direct seed health tests, can be found in Tables 5B of Chapter 5 of the ISTA Rules, or in the *Handbook of Tolerances and Measures of Precision for Seed Testing* by S.R. Miles (*Proceedings of the International Seed Testing Association* 28 (1963) No 3, p 644).

2. Reporting Results

The result of a seed health test should indicate the scientific name of the pathogen detected and the test method used. When reported on an *ISTA International Seed Analysis Certificate*, results are entered under Other Determinations.

Quality Assurance

Critical Control Points

None listed

References

The following reference was extracted from the ISTA Handbook on Seed Health Testing, Working Sheet No. 13, 1964.

Neergaard, P. and Saad, A., (1962): Seed health testing of rice. A contribution to development of laboratory routine test methods. *Indian Phytopathology*. 15, 85-111.

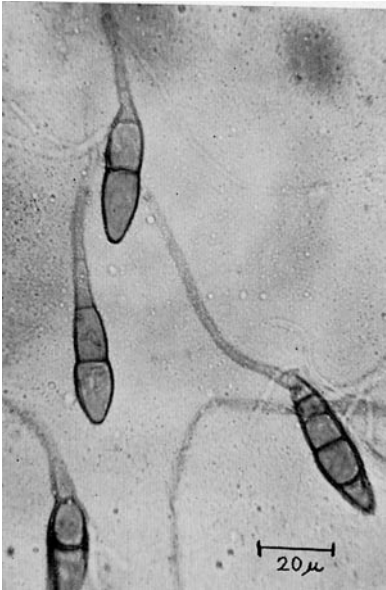


Fig. 1 Conidia

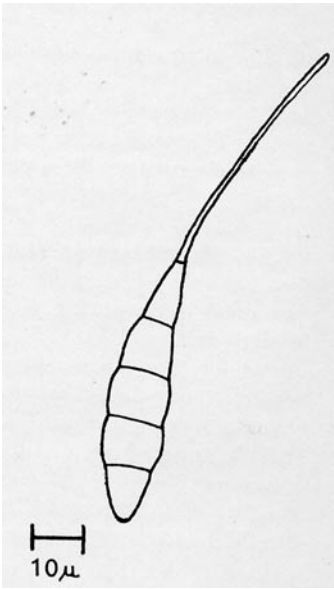


Fig. 2. Conidium

