



ALMA MATER STUDIORUM
UNIVERSITÀ DI BOLOGNA

New Breeding Techniques - an update

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ISTA Seminar: From Biodiversity to Diversification:
resources, tools and technologies to meet new
challenges.

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Gene editing methods

Methods based on hybrid proteins, or protein-RNA complexes, able to target specific DNA regions where they induce mutations

- **MEGANUCLEASES:** enzymes with long target recognition sequences (14-40 nt) and high DNA cleavage specificity, derived from transposon-like elements from mitochondria, chloroplast and bacteria genomes
- **ZFN (Zinc-Finger Nucleases):** DNA binding Zinc-finger motif is associated with the nuclease motif of *FokI*
- **TALEN (Transcription Activator-Like Effector Nucleases):** DNA binding motif of a transcription activator from *Xanthomonas* associated with *FokI*.
- **CRISPR-Cas9 (Clustered-regularly interspaced short palindromic repeats – Crisp associated protein9).**

Nobel Prize for Chemistry (2020)

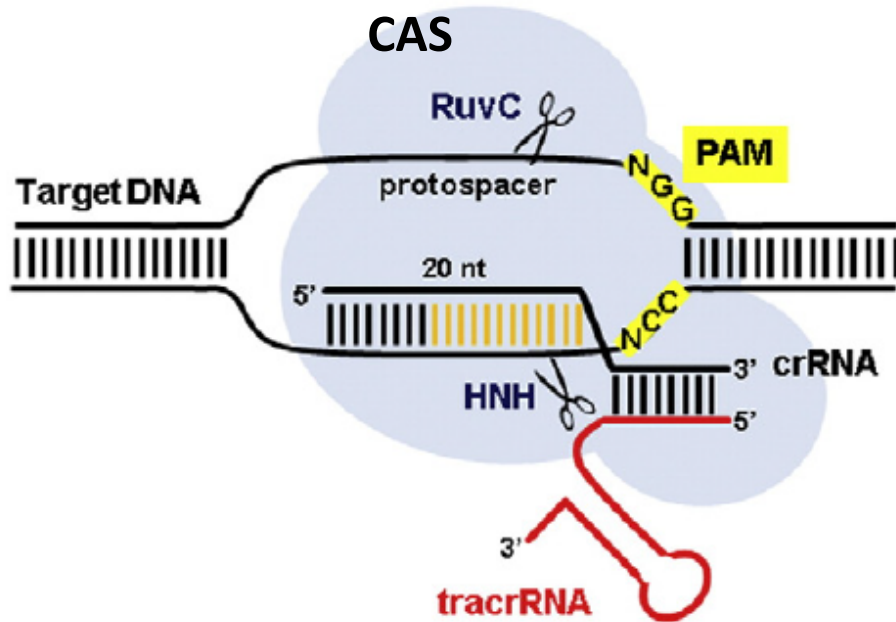


Emmanuelle Carpentier, Umea
University, Svezia
(now at Max Planck Institute for
Infection Biology)

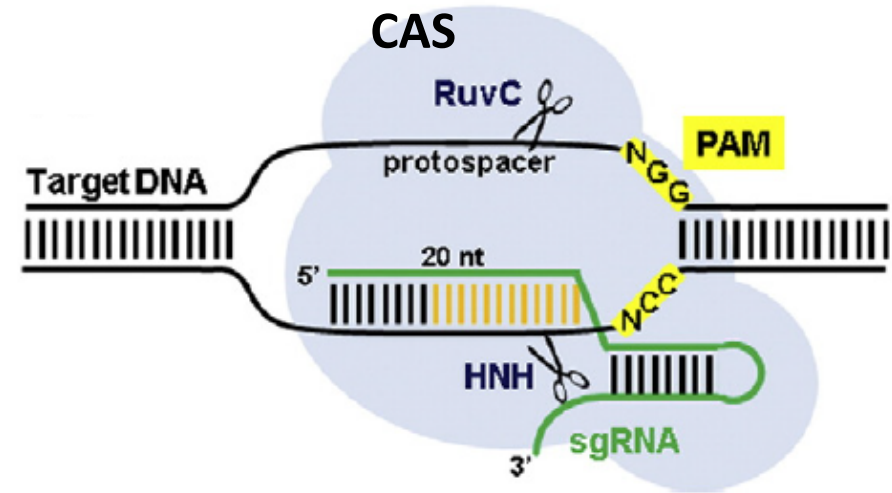


Jennifer Doudna,
University of Berkeley

CRISPR-CAS main components



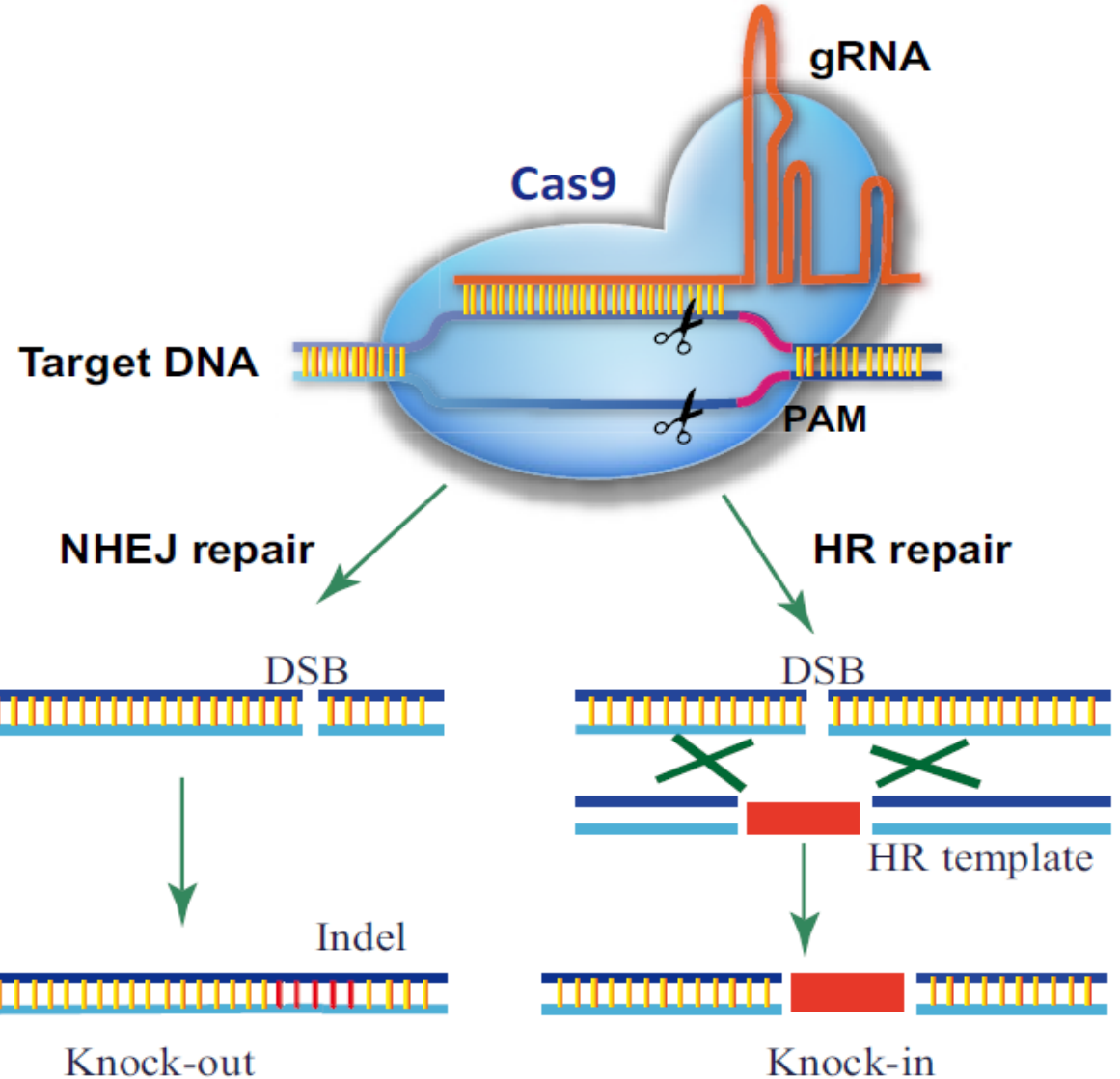
crRNA = CRISPR-RNA
tracrRNA = transactivating crRNA
CAS = CRISPR-Associated nuclease
Native



sgRNA = single guide RNA

Modified as gene
editing tool

Mechanisms of targeted mutagenesis with CRISPR-CAS



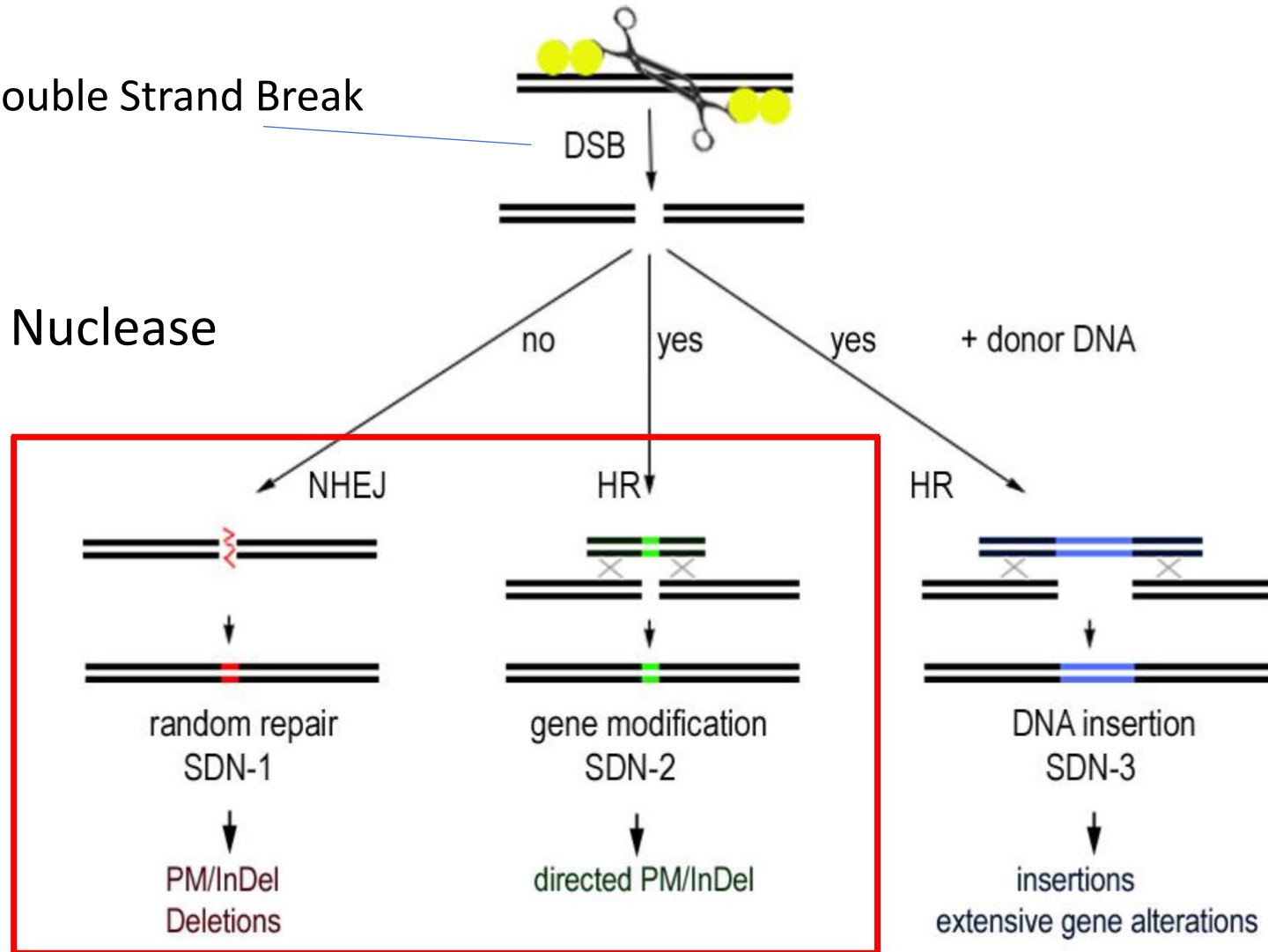
NHEJ (non-homologous end joining)

HR (homology-directed repair)

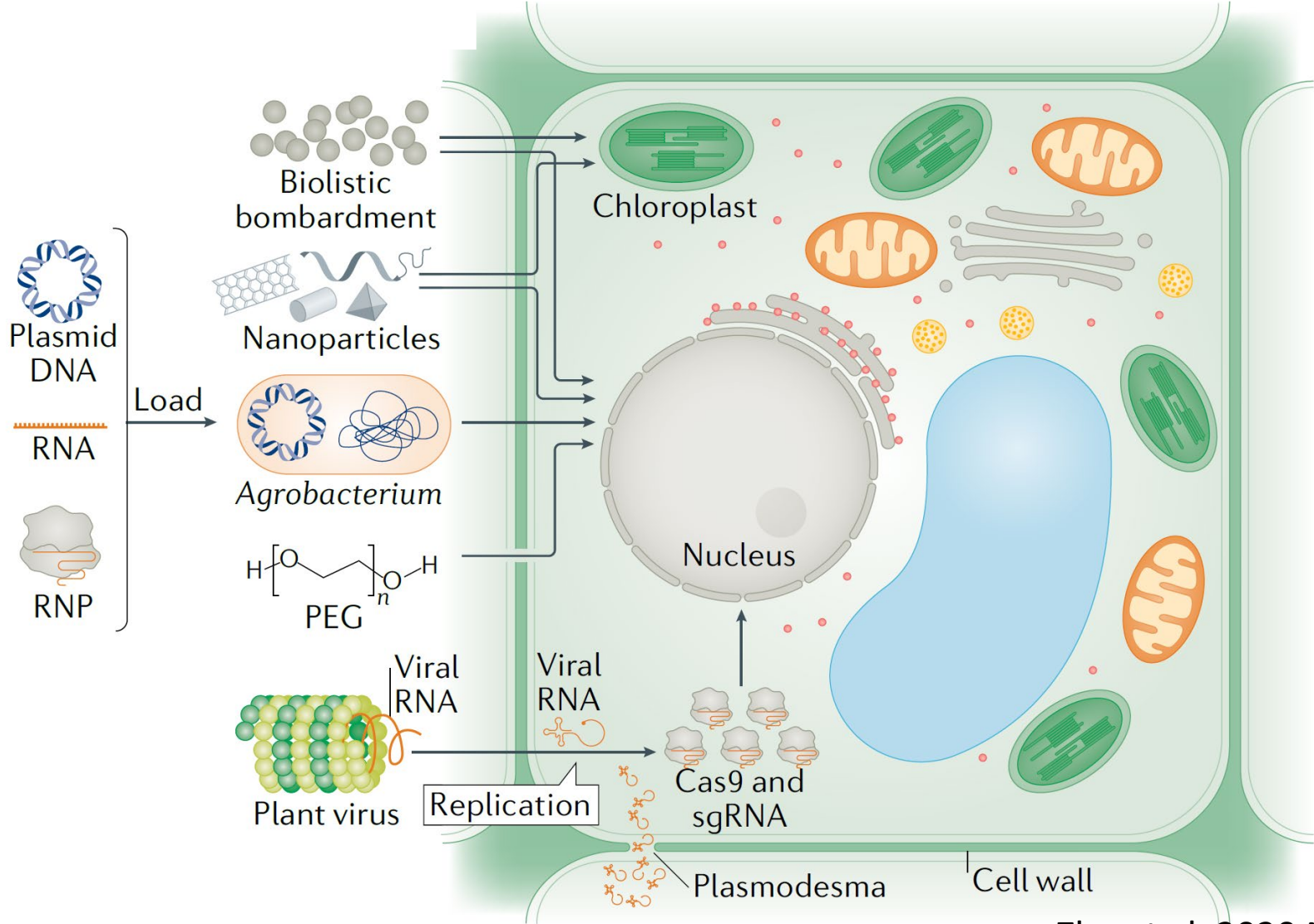
The 'SDN' system to classify edited events

DSB = Double Strand Break

SDN = Site-Directed Nuclease technologies, 1-3



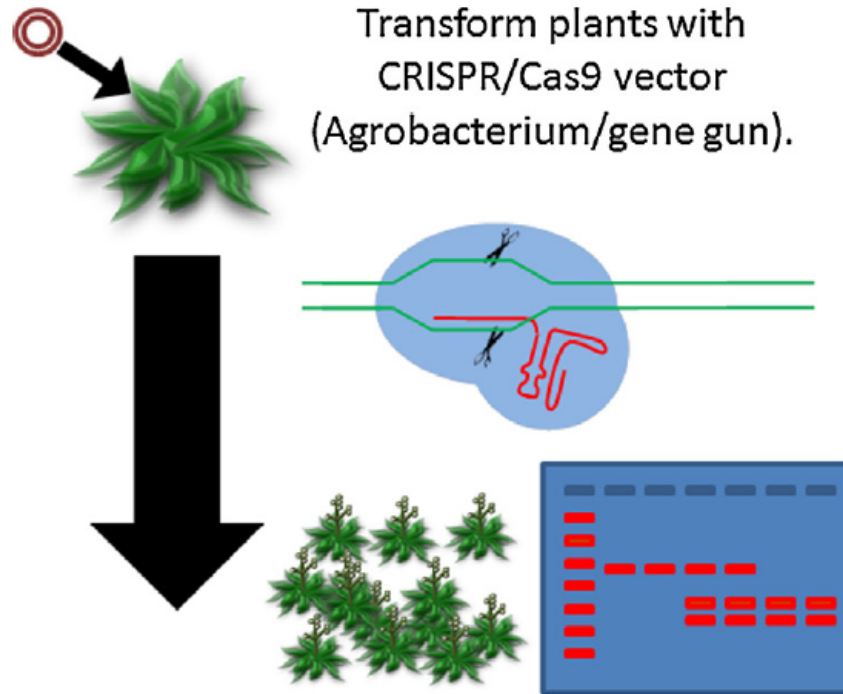
Delivering CRISPR/Cas reagents



Delivering CRISPR/Cas reagents

- Conventional delivery methods are still *Agrobacterium*-mediated transformation and particle bombardment
 - however new methods seem promising
- Particle bombardment (or PEG-mediated transfection) using RNP
 - Boosted with regeneration, or meristem, inducers (*BBM1* and *WUS2*, etc)
- Viral vectors-mediated delivery system
- Nanoparticle-based transformation
- Grafting-based systems

CRISPR-CAS editing traditionally goes through transgenics



Transform plants with
CRISPR/Cas9 vector
(Agrobacterium/gene gun).

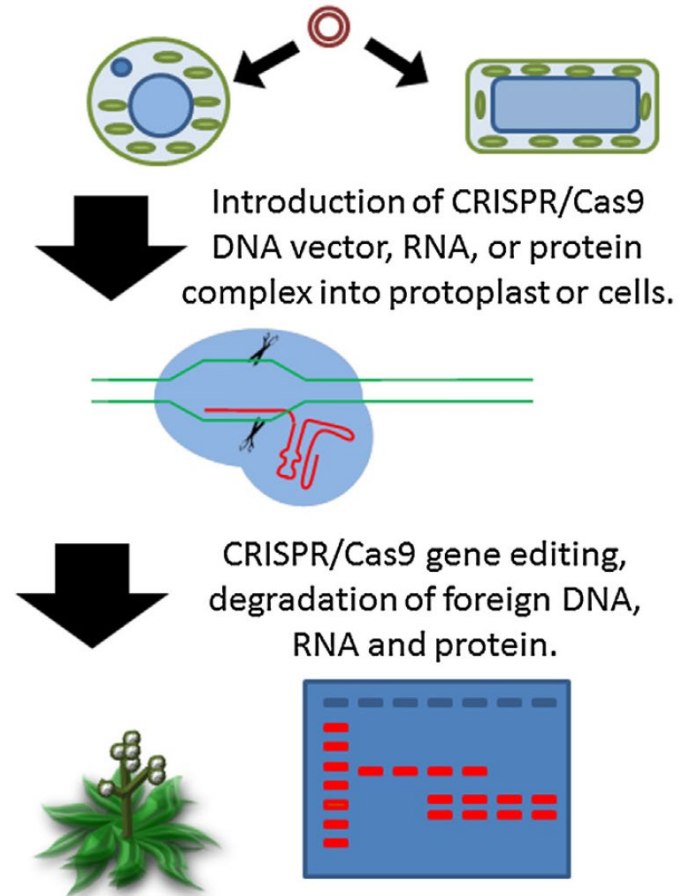
Screen T₀ plants for homozygous mutation
using PCR and restriction digest. Self or
back-cross with wildtype.



**Screen T₁/T₂ plants for mutation
presence and transgene absence**

*T-DNA with gRNA-CAS9 construct
is expected to land on a different
chromosome from the one carrying
the target gene, so it can be
segregated off in subsequent
generations*

RNP-based delivery may produce edits without transgenics

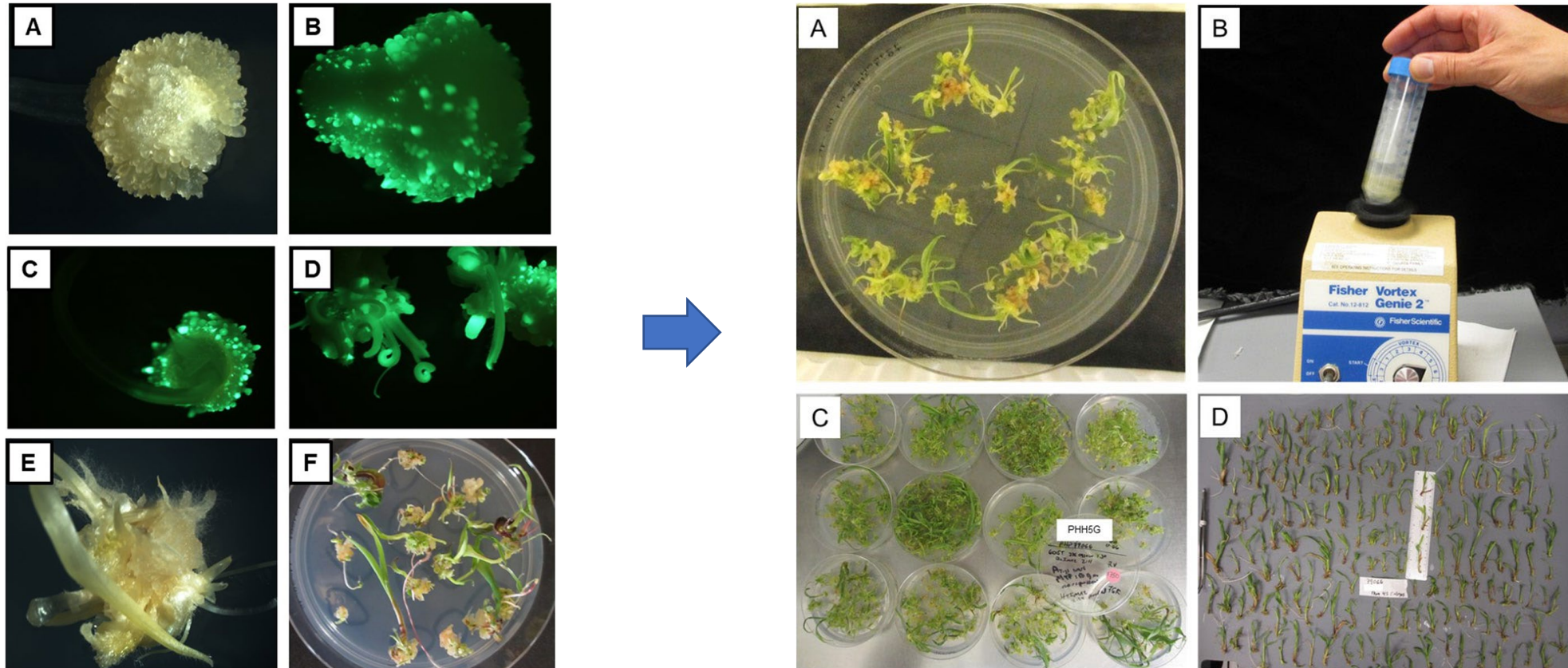


RNP = Ribonucleoprotein =
gRNA + CAS9 (protein or DNA
or RNA)

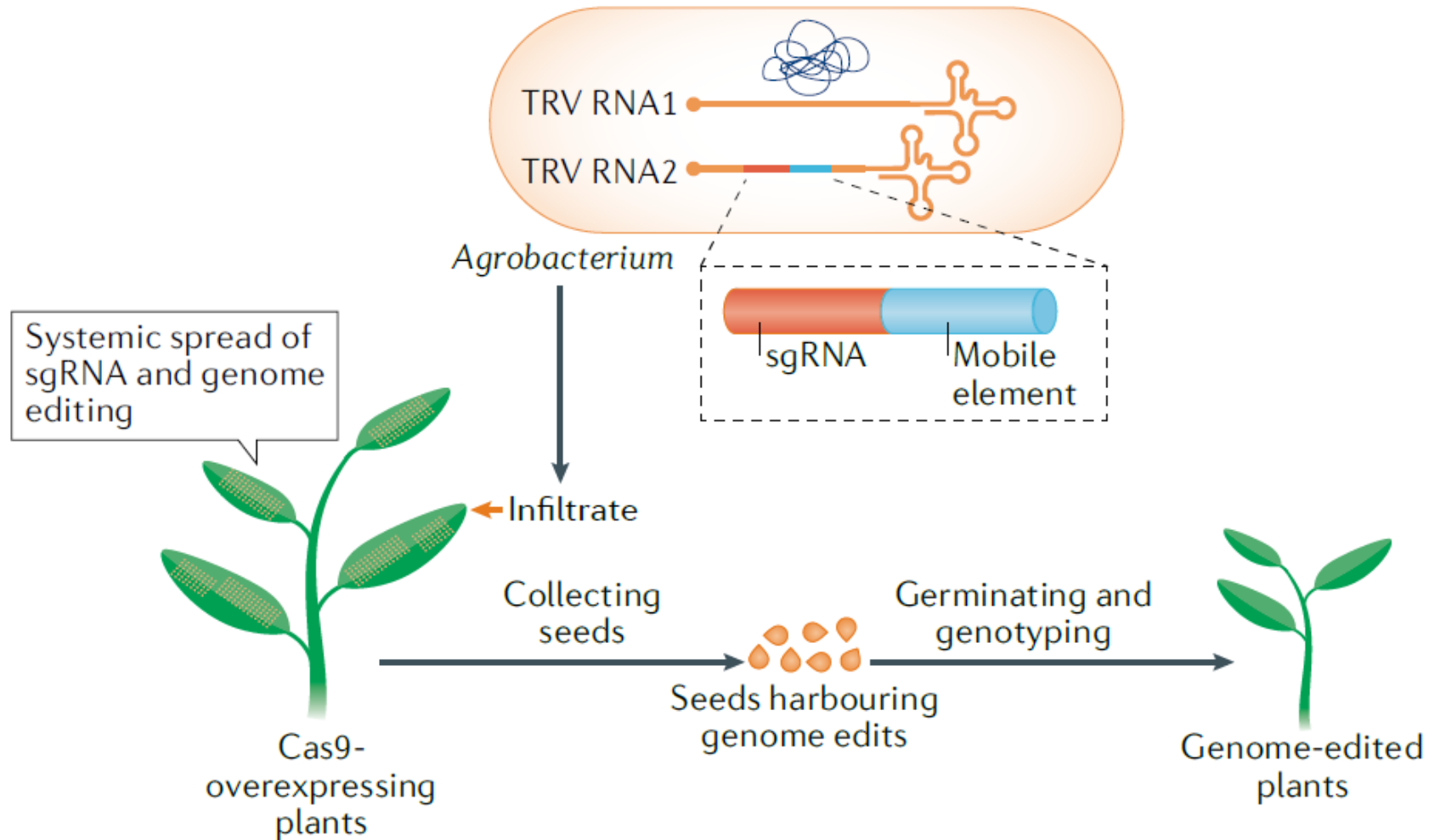
**Regeneration T1/T2 plants and screen
for mutation**

Use of morphogenetic regulators to boost regeneration

BABY BOOM 1 (BBM1) and *WUS2* in co-transformation or assembled in RNP



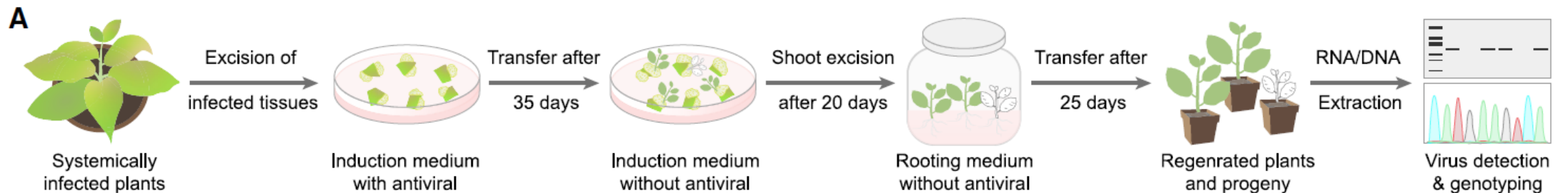
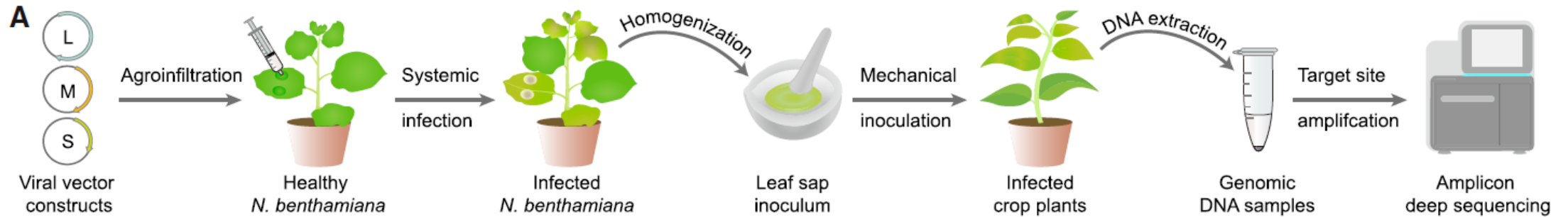
Virus-induced heritable gene editing



Virus-based vectors: miniature cargos for CRISPR machinery

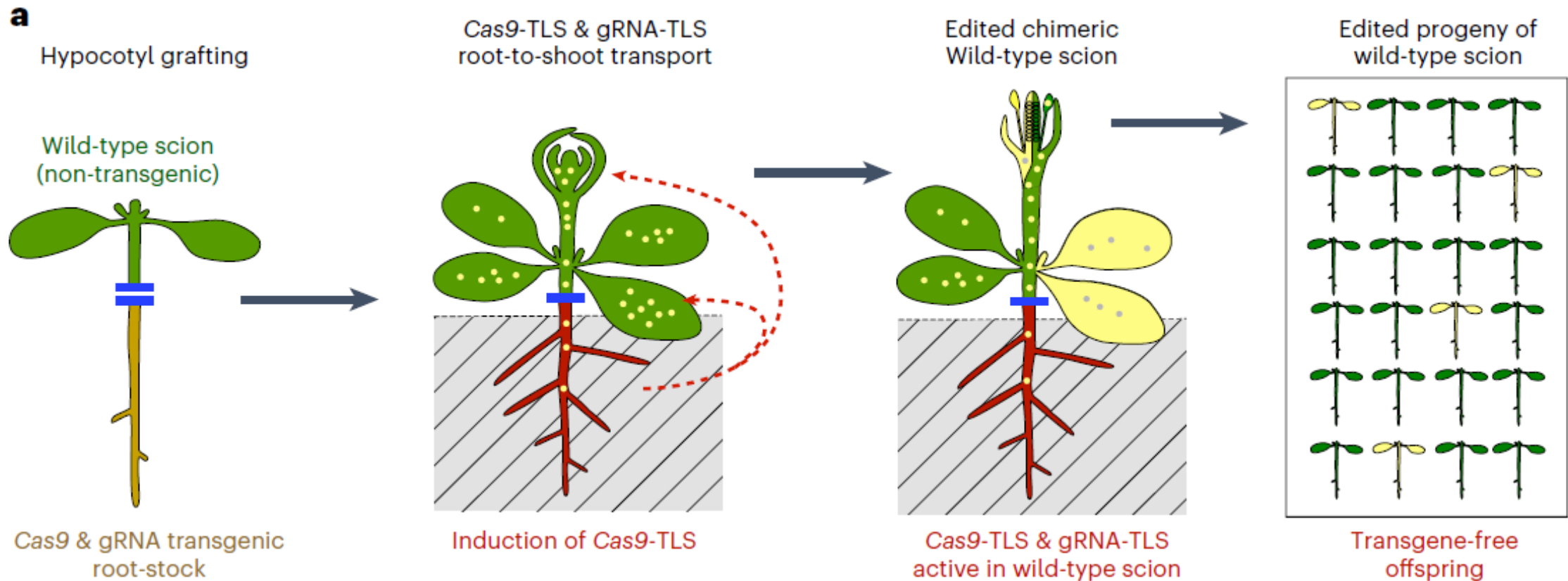
- various virus-based vectors can be utilized to deliver genome-editing reagents without risking the genetic integration of foreign DNA into the plant genome.
- tobacco rattle virus (TRV) and potato virus X (PVX), have been used as carriers for targeted mutagenesis in plants, but owing to their limited cargo capacity, only single guide RNAs (sgRNAs) have been delivered in Cas9-overexpressing (Cas9-OE) transgenic lines.
- Recently, Liu et al. reported a tomato spotted wilt virus (TSWV)-based vector for the delivery of CRISPR/Cas9 and Cas12a machinery in various crops for targeted mutagenesis

Virus-induced heritable gene editing (‘-’RNA virus cargos with both sgRNA and CAS)



CRISPR–Cas9-mediated transgene-free gene editing by grafting

Fusions of Cas9 and guide RNA transcripts to **tRNA-like sequence** motifs that move RNAs from transgenic rootstocks to grafted wild-type shoots (scions)



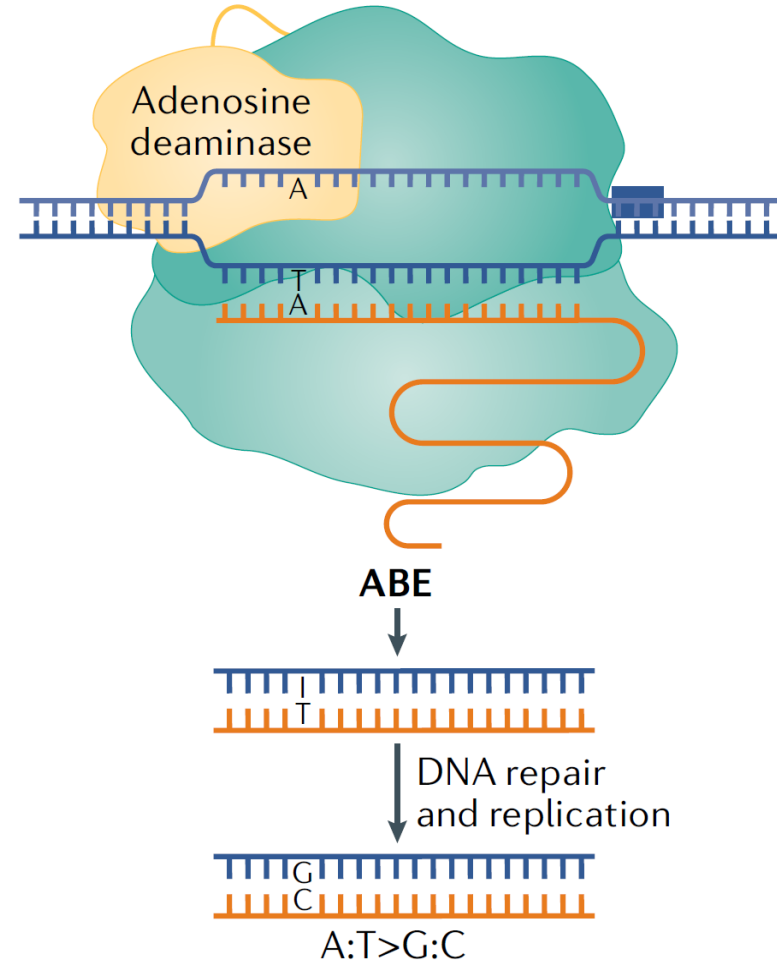
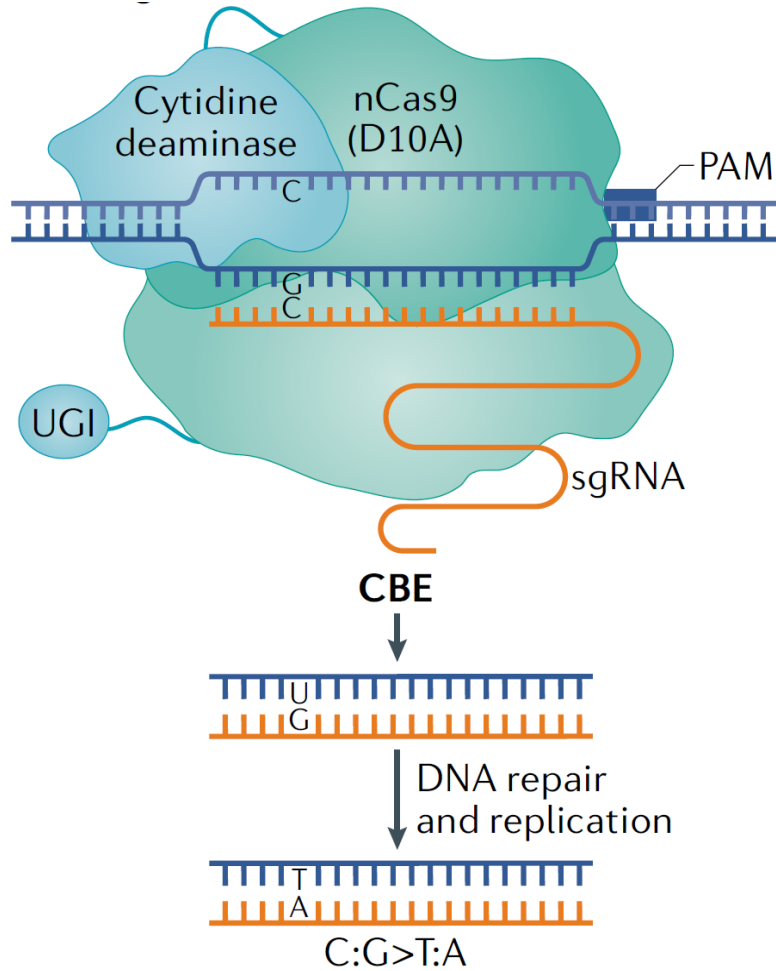
Improving the specificity of genome editing (ie. reduction of off-targets)

- Correct sgRNA design (eg 40-60% GC, and other constraints)
- Chemical modification of sgRNA (eg. Integration of bridges and locks)
- Use of Cas9-gRNA ribonucleoproteins (RNP)s
- Engineered precision variants of Cas9, Cas12a, and deaminases or high-fidelity Cas9 (eg. enhanced specificity SpCas9, eSp-Cas9)

Improving the range of genome edits

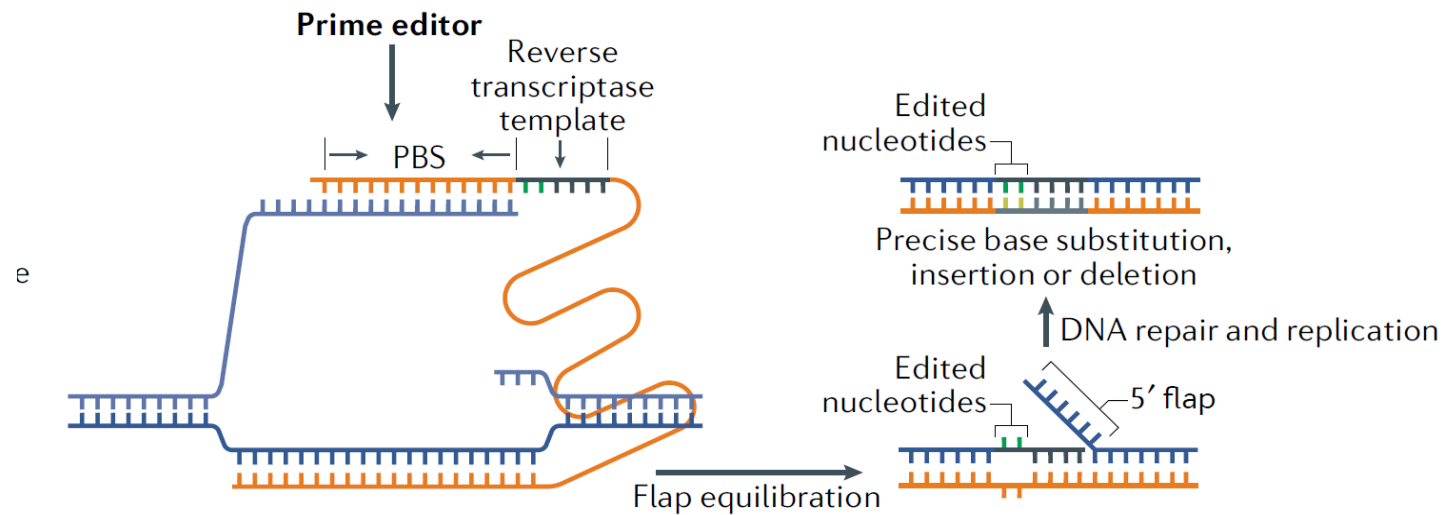
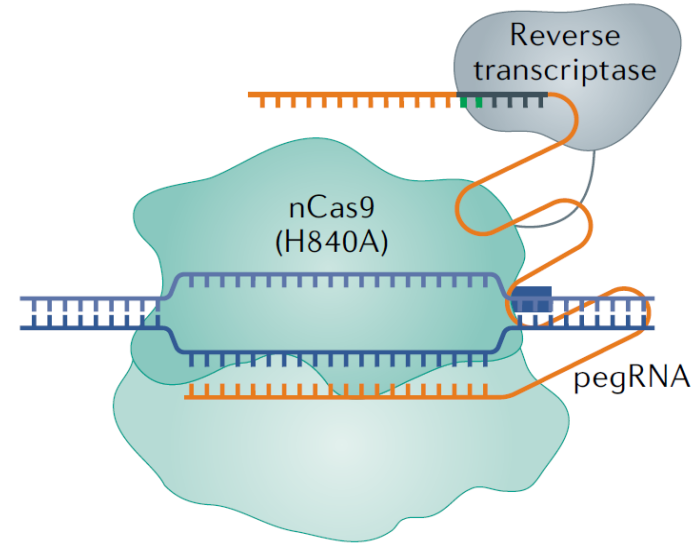
Cytosine and Adenine base editing

Generate base transition!

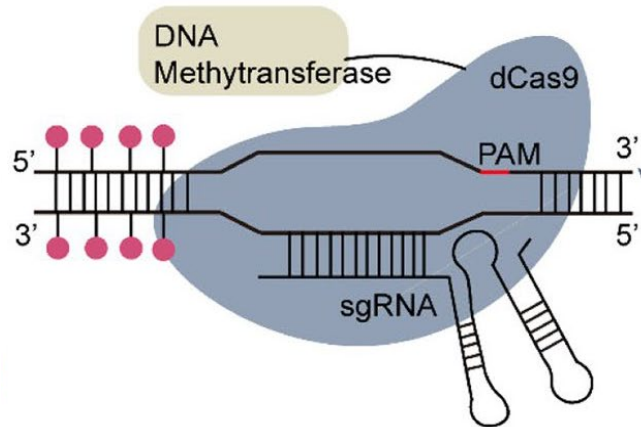


Prime editing

- Can produce all 12 kinds of base substitutions on target (at least in human cells)
- Under optimization in plants

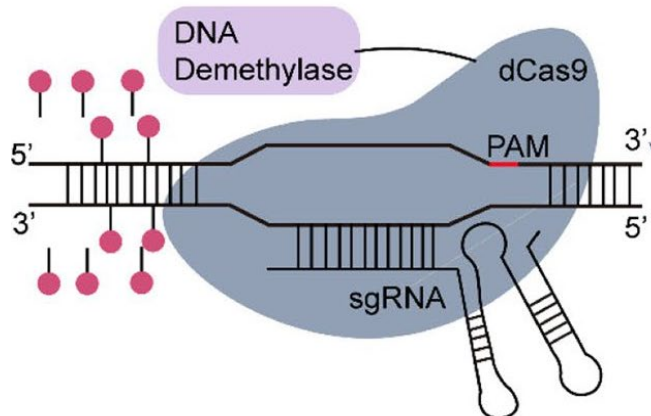


CRISPR/dCas9-based epigenetic modifier



Expression of Demethylase Genes
CsDME, CsDML1, CsDML3, CsDML4

DNA Methylation Level



Expression of Methyltransferase Genes
Expression of Genes Involved in RdDM

DNA Methylation Level

Conclusions

- Gene editing by CRISPR-CAS has already proved to be applicable to crops
- The protocols and molecular components are being further optimized, so scope, efficiency and precision will likely improve strongly in the near future
- The regulation is clearly the current main obstacle to full exploitation of this technology

Gene editing by CRISPR-CAS method

